

## Luffariolides H and J, New Sesterterpenes from a Marine Sponge *Luffariella* Species

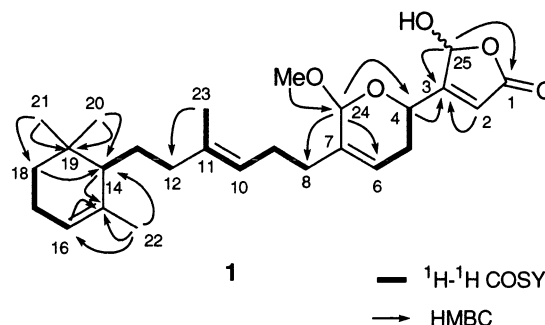
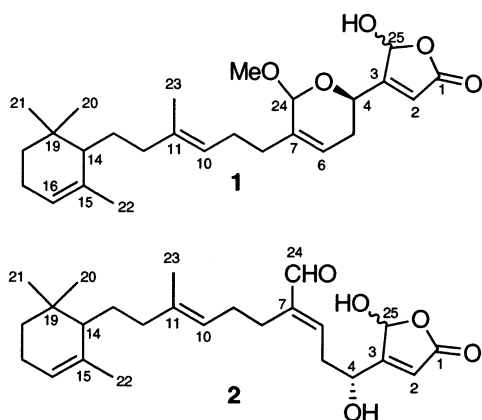
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Two new sesterterpenoids, luffariolides H (**1**) and J (**2**), have been isolated from an Okinawan marine sponge *Luffariella* sp. The structures of **1** and **2** were elucidated on the basis of spectroscopic data.

Marine sponges of the genus *Luffariella* are a rich source of sesterterpenoids with biological activity.<sup>1</sup> In our search for bioactive substances from marine sponges,<sup>2</sup> we have investigated bioactive metabolites from marine sponges of the genus *Luffariella*.<sup>3–6</sup> Our recent study of extracts of an Okinawan *Luffariella* sponge (SS-996) resulted in the isolation of two new sesterterpenoids, luffariolides H (**1**) and J (**2**). Here we describe the isolation and structure elucidation of **1** and **2**.



**Figure 1.** Selected 2D NMR correlations for luffariolide H (**1**).

presence of an epimeric hydroxyl group at C-25, to which a similar phenomenon was reported for manolide.<sup>3</sup> Analysis of 2D NMR data (Figure 1) suggested that the C-1–C-13 part of **1** possessed the same structure as that of manolide. The HMBC correlation for a methoxy signal to C-24 indicated that the methoxy group was attached to C-24. Comparing <sup>13</sup>C NMR data (Table 1) of **1** with those of manolide, differences were observed for the chemical shifts of C-14, C-15, C-16, and C-19. The HMBC spectrum of **1** showed correlations for H<sub>2</sub>-18, H<sub>3</sub>-21, and H<sub>3</sub>-22 to C-14, H-14, H-16, and H<sub>3</sub>-22 to C-15, and H<sub>2</sub>-19, H<sub>3</sub>-20, and H<sub>3</sub>-21 to C-19, indicating the presence of a  $\Delta^{15(16)}$  cyclohexene ring. This assignment was supported by comparison of NMR data for the C-13–C-22 part of **1** with those for the corresponding moiety of cacospongins B and C<sup>10</sup> and luffarin P.<sup>11</sup> Thus, the structure of luffariolide H was elucidated to be **1**.

HRFABMS data [ $m/z$  439.2485 ( $M + Na$ )<sup>+</sup>,  $\Delta +2.4$  mmu] of luffariolide J (**2**) suggested the molecular formula C<sub>25</sub>H<sub>36</sub>O<sub>5</sub>. <sup>1</sup>H and <sup>13</sup>C NMR data (Table 1) of **2** were similar to those of secomanolide<sup>4</sup> and luffariellin B,<sup>5</sup> except for those of C-14, C-15, C-16, and C-19. The presence of a  $\Delta^{15(16)}$  cyclohexene ring in **2** was implied by 2D NMR data and comparison of the <sup>13</sup>C chemical shifts with those of **1**. Thus, the structure of luffariolide J was assigned as **2**.

Absolute configurations at C-4 of luffariolides H (**1**) and J (**2**) were considered to be *R* judging from co-isolation of 24-*O*-methylmanolide<sup>7</sup> and secomanolide<sup>8</sup> with the 4*R*-configuration from this sponge, while those of C-14 were not defined.

Several sesterterpenoids have been isolated from *Luffariella* sponges.<sup>7–11</sup> Luffariolides H (**1**) and J (**2**) exhibited cytotoxicity against murine lymphoma L1210 (IC<sub>50</sub>, 3.2 and 4.5  $\mu$ g/mL, respectively) and epidermoid carcinoma KB cells (IC<sub>50</sub>, 7.5 and 8.2  $\mu$ g/mL, respectively) in vitro. Luffariolides H (**1**) and J (**2**) showed antimicrobial

The sponge *Luffariella* sp. (0.3 kg, wet weight) was extracted with MeOH, and the extracts were partitioned between EtOAc and water. The EtOAc-soluble materials were subjected to silica gel column chromatography (CHCl<sub>3</sub>/MeOH) followed by reversed-phase HPLC (MeOH/H<sub>2</sub>O/AcOH and then CH<sub>3</sub>CN/NH<sub>4</sub>Cl(aq)) to afford luffariolides H (**1**, 4.0 mg, 0.005%, wet weight) and J (**2**, 3.9 mg, 0.005%) together with known related sesterterpenoids, 24-*O*-methylmanolide,<sup>7,8</sup> secomanolide,<sup>8</sup> and luffariellins A<sup>9</sup> and B.<sup>9</sup>

Luffariolide H (**1**) showed a molecular ion peak at  $m/z$  430 in the EIMS spectrum, and the molecular formula, C<sub>26</sub>H<sub>38</sub>O<sub>5</sub>, was confirmed by HREIMS ( $m/z$  430.2698  $M^+$ ,  $\Delta -2.1$  mmu). IR absorptions at 3375 and 1760  $\text{cm}^{-1}$  of **1** were attributed to a hydroxyl group and an unsaturated  $\gamma$ -lactone moiety, respectively. <sup>1</sup>H and <sup>13</sup>C NMR data (Table 1) were reminiscent of those of manolide,<sup>7</sup> except for the presence of a methoxy group. The signals for H-2, H-4, H-24, and H-25 were observed as doublets due to the

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**Table 1.**  $^1\text{H}$  and  $^{13}\text{C}$  NMR Data of Luffariolides H (**1**) and J (**2**) in  $\text{CDCl}_3$ 

positn	<b>1</b>				<b>2</b>								
	$\delta_{\text{H}}^a$		$\delta_{\text{C}}^b$		$\delta_{\text{H}}^a$		$\delta_{\text{C}}^b$						
1			171.8				170.4	171.2					
2	6.05	s	6.13	s	117.5	118.4	6.03	s	6.11	s	118.3	118.4	d
3			167.2		167.5						169.3		s
4	4.76	m	4.82	m	62.2	63.1	4.76	m	4.83	m	66.4	66.9	d
5	2.28 <sup>a</sup>	m			29.1		2.82 <sup>a</sup>	m			24.4		t
6	5.68	brs			120.6	120.7	6.55	m			145.6	145.8	d
7			136.4								148.3	148.8	s
8	2.09 <sup>a</sup>	m			32.5	32.6	2.03 <sup>a</sup>	m			34.3	34.7	t
9	2.34 <sup>a</sup>	dd, 5.8, 5.9			26.1		2.27 <sup>a</sup>	m			26.8		t
10	5.11	t, 5.8			123.3		5.08	m			122.7		d
11			136.7								136.8		s
12	2.10 <sup>a</sup>	m			40.5		1.95	m			40.5		t
13	1.35	m	1.52	m	29.9		1.32	m	1.47	m	29.8		t
14	1.42	t, 3.6			49.1		1.38	m			49.0		d
15			136.9								137.3		s
16	5.29	brs			120.6		5.26	m			120.0		d
17	1.95 <sup>a</sup>	m			23.1		1.93 <sup>a</sup>	m			23.0		t
18	1.14 <sup>a</sup>	t, 4.5			31.8		1.11 <sup>a</sup>	m			31.6		t
19			28.7								32.5		s
20	0.87 <sup>b</sup>	s			27.4		0.84 <sup>b</sup>	s			27.4		q
21	0.92 <sup>b</sup>	s			27.5		0.95 <sup>b</sup>	s			27.5		q
22	1.68 <sup>b</sup>	s			23.5		1.65 <sup>b</sup>	s			23.5		q
23	1.61 <sup>b</sup>	s			16.1		1.56 <sup>b</sup>	s			16.1		q
24	4.79	s	4.81	s	98.3	98.5	9.37	s			195.3		d
25	6.10	s	6.27	s	97.1	97.6	6.10	s	6.26	s	98.0	98.3	d
OCH <sub>3</sub>	3.45 <sup>b</sup>	s			55.8			q					q

<sup>a</sup> 2H. <sup>b</sup> 3H.

activity against *Staphylococcus aureus* (MIC, 16.7 and 33.3  $\mu\text{g/mL}$ , respectively), *Bacillus subtilis* (MIC, both 8.4  $\mu\text{g/mL}$ ), and *Mycrococcus luteus* (MIC, both 8.4  $\mu\text{g/mL}$ ).

### Experimental Section

**General Experimental Procedures.** The IR and UV spectra were recorded on a JASCO FT/IR-5300 and a Shimadzu UV-1600PC spectrophotometer, respectively.  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectra were recorded on a Bruker AMX-600 spectrometer. EI mass spectra were recorded on a JEOL FABmate spectrometer at 70 eV. FAB mass spectrum was obtained on a JEOL HX-110 spectrometer using nitrobenzyl alcohol as a matrix.

**Animal Material.** The sponge (order Dictyoceratida, family Thorectidae) was collected off Nakijin, Okinawa, and kept frozen until used. The sponge had a medium brown finely conulose surface skin with an orange-brown interior after preservation. The sponge was yellow-brown when alive. The sponge is very firm, moderately compressible, and springy. The thin conulose skin is 70  $\mu\text{m}$  thick and consists of thickened mesohyl and a thin layer of sand grains. The primary fibers are thick, striated, and centrally cored with sand grains. They are 200  $\mu\text{m}$  thick with mesh spaces of up to 1000  $\mu\text{m}$ . They can be fasciculate toward the surface. The secondary fibers are striated, uncored, and 40–90  $\mu\text{m}$  thick with mesh spaces up to 375  $\mu\text{m}$ . The tertiary fibers are 10–13  $\mu\text{m}$  thick with small mesh spaces.

**Extraction and Isolation.** The sponge (0.3 kg, wet weight) was extracted with MeOH (600 mL  $\times$  2), and the extract was partitioned between EtOAc (500 mL  $\times$  3) and H<sub>2</sub>O (500 mL). Parts (1.1 g) of the EtOAc-soluble materials (1.67 g) were subjected to a silica gel column ( $\text{CHCl}_3/\text{MeOH}$ ). The fraction eluted with  $\text{CHCl}_3/\text{MeOH}$  (98:2) was purified by reversed-phase HPLC [Luna Phenyl-hexyl, 5  $\mu\text{m}$ , Phenomenex, 10  $\times$  250 mm; eluent, MeOH/H<sub>2</sub>O/AcOH, 85:15:0.1; flow rate, 2.5 mL/min; UV detection at 230 nm] and then C<sub>18</sub> HPLC [Wakosil-II5C18 RS, Wako Pure Chemical Ind., Ltd., 10  $\times$  250 mm; eluent, CH<sub>3</sub>CN/5 mM NH<sub>4</sub>Cl(aq), 75:25; flow rate, 2.5 mL/min; UV detection at 230 nm] to afford luffariolide H (**1**, 4.0 mg, 0.005%, wet weight), 24-O-methylmanoalide (0.002%), and

luffariellin A (0.003%). The fraction eluted with  $\text{CHCl}_3/\text{MeOH}$  (95:5) in the first silica gel column was subjected to C<sub>18</sub> HPLC [Wakosil-II5C18 RS, 10  $\times$  250 mm; eluent, CH<sub>3</sub>CN/5 mM NH<sub>4</sub>Cl(aq), 60:40; flow rate, 2.5 mL/min; UV detection at 230 nm] to give luffariolide J (**2**, 3.9 mg, 0.005%), secmanoalide (0.002%), and luffariellin B (0.003%).

**Luffariolide H (1):** colorless oil; UV (MeOH)  $\lambda_{\text{max}}$  230 nm (sh); IR (KBr)  $\nu_{\text{max}}$  3375 and 1760  $\text{cm}^{-1}$ ;  $^1\text{H}$  and  $^{13}\text{C}$  NMR (see Table 1); EIMS  $m/z$  398 [M – MeOH]<sup>+</sup> and 430 [M]<sup>+</sup>; HREIMS  $m/z$  430.2698 [M]<sup>+</sup> (calcd for C<sub>26</sub>H<sub>38</sub>O<sub>5</sub>, 430.2719).

**Luffariolide J (2):** colorless oil; UV (MeOH)  $\lambda_{\text{max}}$  230 nm (sh); IR (KBr)  $\nu_{\text{max}}$  3355, 1775, and 1680  $\text{cm}^{-1}$ ;  $^1\text{H}$  and  $^{13}\text{C}$  NMR (see Table 1); FABMS  $m/z$  439 [M + Na]<sup>+</sup>; HRFABMS  $m/z$  439.2485 [M + Na]<sup>+</sup> (calcd for C<sub>25</sub>H<sub>36</sub>O<sub>5</sub>Na, 439.2509).

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